1: marinomycin A ( $trans \Delta^{8,9}$ ,  $trans \Delta^{8',9'}$ )

2: marinomycin B ( $cis \Delta^{8,9}$ ,  $cis \Delta^{8',9'}$ ) 3: marinomycin C ( $trans \Delta^{8,9}$ ,  $cis \Delta^{8',9'}$ )

Scheme 1. Structures of marinomycins A-C (1-3).

Natural Product Synthesis

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## Total Synthesis of Marinomycins A-C\*\*

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Marinomycins A–C (1–3, Scheme 1) are recently discovered natural products with imposing molecular architectures and impressive biological properties. [1] Isolated from actinomycete *Marinispora* strain CNQ-140 cultured from a sediment collected from the bottom of the ocean offshore of La Jolla, California (USA), by Fenical and co-workers, [1] these novel compounds exhibit significant antibiotic activities (minimum inhibitory concentration, MIC = 0.1–0.6 μm) against methicil-lin-resistant *Staphylococcus aureus* (MRSA) and vancomycin-resistant *Enterococcus faecium* (VREF), and inhibit cancer cell proliferation against the National Cancer Institute's 60 cancer cell line panel (LC<sub>50</sub> = 0.2–2.7 μm). In particular, these marinomycins 1–3 showed potent and selective cytotoxicities against six of the eight melanoma cell lines of that panel. [1] Their novel and sensitive polyunsa-

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Supporting information for this article is available on the WWW under http://www.angewandte.org or from the author.

turated structures coupled with their potentially useful biological activities prompted our interest in these molecules. Herein, we report the total synthesis of marinomycins A–C (1–3) and of two of their hitherto unknown monomeric homologues, mono-marinomycin A (m-1) and *iso*-mono-marinomycin A (m-2, Scheme 7).

Given that 1, the most abundant of the marinomycins, is photolytically convertible to a mixture of all three (i.e. 1, 2, and 3),<sup>[1]</sup> a total synthesis of marinomycin A (1) would constitute syntheses of 2 and 3 as well. Scheme 2 depicts our

**Scheme 2.** Retrosynthetic analysis of marinomycin A (1). HWE = Horner–Wadsworth–Emmons, TBS = *tert*-butyldimethylsilyl, TES = triethylsilyl.

6

**OTES** 

BnÓ

5

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original retrosynthetic analysis of marinomycin A (1). The symmetrical structure of the molecule renders it suitable for several retrosynthetic disconnections; the one shown in Scheme 2 was chosen to highlight and test the Suzuki reaction as a means to construct large and complex macrocycles. <sup>[2]</sup> The appropriately functionalized vinyl boronic acid vinyl bromide 4 needed for the originally intended dimerization was traced to the three key building blocks ketophosphonate 5, aldehyde 6, and carboxylic acid 7, through the indicated Horner–Wadworth–Emmons (HWE) olefination and Mitsunobu reactions. It was expected that dimer versus monomer formation in the cyclization process would be subject to concentration conditions.

The required building blocks **5–7** were synthesized in their enantiomerically pure forms as summarized in Schemes 3–5. Starting with the construction of ketophosphonate **5** (Scheme 3), the enantiomerically pure epoxide **8**<sup>[3]</sup> was regioselectively opened in the presence of BF<sub>3</sub>·OEt<sub>2</sub> with the lithium reagent derived from propargylic ether **9**<sup>[4]</sup> and *n*BuLi at –78°C, leading to the corresponding secondary alcohol (89% yield), whose silylation (TBSCl, 96% yield) and de-*p*-methoxybenzylation (DDQ, 79% yield) afforded the chain-extended propargylic alcohol **10**. Exposure of the latter compound to Red-Al, followed by addition of NIS, furnished the corresponding hydroxy vinyl iodide (66% yield), whose temporary silylation (TMSCl) and subsequent coupling with ZnMe<sub>2</sub> in the presence of [Pd(dppf)Cl<sub>2</sub>] (cat.)

Scheme 3. Preparation of ketophosphonate 5. Reagents and conditions: a) 9 (2.0 equiv), nBuLi (2.5 m in hexanes, 2.0 equiv), THF, -78 °C, 45 min; then BF<sub>3</sub>·OEt<sub>2</sub> (1.2 equiv), **8** in THF, -78 °C, 4 h, 89%; b) TBSCl (1.2 equiv), imid. (3.0 equiv), DMF, 25 °C, 6 h, 96%; c) DDQ (1.5 equiv), CH<sub>2</sub>Cl<sub>2</sub>, pH 7 phosphate buffer, 25 °C, 3 h, 79%; d) Red-Al (3.33 M in toluene, 1.7 equiv), THF, 25 °C, 45 min; then NIS (1.8 equiv), THF, -78 °C, 30 min, 66%; e) TMSCI (2.0 equiv), Et<sub>3</sub>N (5.0 equiv), 25 °C, 2 h; then [Pd(dppf)Cl<sub>2</sub>] (0.05 equiv), Me<sub>2</sub>Zn (2.0 equiv), THF, 65 °C, 12 h; then K<sub>2</sub>CO<sub>3</sub> (0.1 equiv), MeOH, 25 °C, 4 h, 81 %; f) NaH (1.6 equiv), BnBr (1.7 equiv), nBu<sub>4</sub>NI (0.1 equiv), THF, 25 °C, 6 h, 88%; g) HF-py, THF, 0°C, 3 h, 77%; h) PCC (2.0 equiv), NaHCO $_3$  (0.5 equiv), 25°C, 3 h, 73%; i) CH<sub>3</sub>P(O) (OMe)<sub>2</sub> (4.0 equiv), nBuLi (2.5 м in hexanes, 4.0 equiv), THF, -78°C, 2 h; then 13 in THF, -78°C, 2 h; j) PDC (2.1 equiv), 4-Å M.S., DMF, 25 °C, 12 h, 64% over two steps; k) HF (48% aq), MeCN, 25 °C, 3 h, 92%. Bn = benzyl, DDQ = 2,3-dichloro-5,6-dicyano-1,4-benzoquinone, dppf= bis (diphenylphosphino) ferrocene, DMF = N, N-dimethylformamide, imid. = imidazole, PMB = p-methoxybenzyl, M.S. = molecular sieves, NIS = N-iodosuccinimide, PCC = pyridinium chlorochromate, PDC = pyridinium dichromate, py = pyridine, THF = tetrahydrofuran, TMS = trimethylsilyl.

gave, upon basic (K<sub>2</sub>CO<sub>3</sub>, MeOH) workup, the corresponding primary allylic alcohol (81 % yield). Benzylation of the latter compound (88 % yield) followed by desilylation (HF·py, 77 % yield) and oxidation with PCC afforded aldehyde **13** (73 % yield). Reaction of aldehyde **13** with the lithium species derived from dimethyl methyl phosphonate and *n*BuLi followed by oxidation (PDC) of the resulting epimeric mixture of alcohols led to ketophosphonate **14** in 64 % overall yield (two steps). Finally, fluoride-induced (aq HF/MeCN) desilylation of **14** gave the desired hydroxy ketophosphonate **5** in 92 % yield.

The synthesis of fragment 6 began with the readily available aldehyde  $15^{[5]}$  and involved two iterations of Brown allylation<sup>[6]</sup> [(+)-(ipc)<sub>2</sub>B-allyl]/ozonolysis (PPh<sub>3</sub>) sequences with appropriate protections of the resulting secondary alcohols (Scheme 4). Proceeding through inter-

**Scheme 4.** Preparation of aldehyde **6.** Reagents and conditions: a) (-)-B(lpc)<sub>2</sub>OMe (1.7 equiv), allylMgBr (1.0 m in Et<sub>2</sub>O, 1.7 equiv), Et<sub>2</sub>O,  $-78 \rightarrow 25$  °C, 2 h; then **15** in Et<sub>2</sub>O, -78 °C, 4 h,  $-78 \rightarrow 25$  °C, 1 h, 95% (> 40:1 d.r.); b) TESCl (1.8 equiv), imid. (4.0 equiv), DMF, 25 °C, 4 h; c) O<sub>3</sub>, CH<sub>2</sub>Cl<sub>2</sub>, -78 °C, 4 h; then PPh<sub>3</sub> (1.3 equiv), -78 °C, 2 h,  $-78 \rightarrow 25$  °C, 1 h, 64% over two steps; d) (-)-B(lpc)<sub>2</sub>OMe (1.7 equiv), allylMgBr (1.0 m in Et<sub>2</sub>O, 1.7 equiv), Et<sub>2</sub>O,  $-78 \rightarrow 25$  °C, 2 h; then **17** in Et<sub>2</sub>O, -78 °C, 4 h,  $-78 \rightarrow 25$  °C, 1 h, 85% (> 40:1 d.r.); e) TBSCl (2.0 equiv), imid. (5.0 equiv), DMF, 25 °C, 4 h; f) O<sub>3</sub>, CH<sub>2</sub>Cl<sub>2</sub>, -78 °C, 4 h; then PPh<sub>3</sub> (1.3 equiv), -78 °C, 2 h,  $-78 \rightarrow 25$  °C, 1 h, 91% over two steps. lpc = isopinocampheyl.

mediates **16–18**, this sequence afforded the desired compound **6** in good overall yield (47 % from **15**, six steps) and with high diastereoselectivity (>40:1 d.r.).

The dienyl bromide carboxylic acid **7** was synthesized from known acetonide acetylene **19**<sup>[7]</sup> (Scheme 5). Thus, **19** was reacted with the adduct of BH<sub>3</sub>·THF with 2,5-dimethylhexa-2,4-diene, <sup>[8]</sup> and the resulting borane (87% yield) was coupled with commercially available TMS vinyl bromide **20** in the presence of [Pd(PPh<sub>3</sub>)<sub>4</sub>] (cat.) and Cs<sub>2</sub>CO<sub>3</sub> to afford TMS diene **21** (89% yield). Exposure of the latter compound, **21**, to NBS gave bromide **22** (88% yield), which was converted into acetoxy carboxylic acid **7** through saponification (KOH, 87% yield) and acetylation (Ac<sub>2</sub>O, Mg(ClO<sub>4</sub>)<sub>2</sub>; <sup>[9]</sup> 96% yield).

The assembly of fragments **5–7** and elaboration of the growing molecule to the targeted enyne bromide **28** is shown in Scheme 6. Thus, coupling of ketophosphonate **5** and aldehyde **6** proceeded smoothly under the influence of  $Ba(OH)_2$  to afford the enone in 95 % yield. Hydroxy-directed reduction of this enone with  $Et_2BOMe$  and  $NaBH_4^{[10]}$  at

Scheme 5. Preparation of aryl diene fragment 7. Reagents and conditions: a) 2,5dimethylhexa-2,4-diene (5.5 equiv), BH3·THF (2.5 equiv), THF, 0°C, 3 h; then 19,  $0\,^{\circ}\text{C},\,1.5\text{ h};$  then  $H_2O,\,0{\to}25\,^{\circ}\text{C},\,1\text{ h};$  then  $(\text{CH}_2\text{O})_{\textit{n}},\,25\,^{\circ}\text{C},\,1\text{ h};$  then pinacol (2.0 equiv), 25 °C, 24 h, 87%; b) [Pd(PPh<sub>3</sub>)<sub>4</sub>] (0.1 equiv), Cs<sub>2</sub>CO<sub>3</sub> (10 equiv), 20 (1.3 equiv), THF/H<sub>2</sub>O (2:1), 55 °C, 1 h, 89%; c) NBS (1.2 equiv), MeCN, 25 °C, 15 min, 88%; d) KOH (5.0 equiv), THF/H<sub>2</sub>O (1:1), 55°C, 18 h, 87%; e) Mg(ClO<sub>4</sub>)<sub>2</sub> (0.03 equiv),  $Ac_2O$  (1.1 equiv), 25 °C, 48 h, 96%. NBS = N-bromosuccinimide.

-78 °C resulted in the exclusive formation of the corresponding allylic alcohol (89% yield), whose silvlation (TBSCl) led to the fully protected hexaol 23 (89% yield). The benzyl group was then cleaved from the terminal oxygen atom of this compound, 23, with Ca in liquid NH<sub>3</sub> (75% yield), and the resulting primary alcohol was oxidized with DMP to furnish aldehyde 25 (87% yield). Enyne 26 was then generated from aldehyde 25 upon acetylene installment (TMSCHN<sub>2</sub>-LDA;<sup>[11]</sup> 85% yield) and selective desilylation (TES) with PPTS in ethanol (77% yield). Accompanied by inversion of configuration at C25 (marinomycin numbering scheme), the Mitsunobu reaction between hydroxy compound 26 and carboxylic acid 7 (DEAD, PPh<sub>3</sub>) proceeded in 93 % yield to afford ester 27. Exchanging the Ac groups for a TIPS group (K<sub>2</sub>CO<sub>3</sub>, MeOH; TIPSOTf, 2,6-lut.) then furnished the targeted bromo enyne 28 in excellent yield (92 % overall). This latter switch of protecting groups was necessary because of difficulties encountered in the subsequent hydroboration and Suzuki coupling steps using the Ac-protected and free phenol carboxylic acid variants of 28.

The stage was now set for the anticipated Suzuki dimerization/cyclization in the hope of reaching marinomycins A-C (see Scheme 7). Thus, the required boronic acid 4 was generated from enyne 28 by reaction with catecholborane and catalytic amounts of dicyclohexylborane (THF, 25°C; then H<sub>2</sub>O) and exposed to the action of TlOEt (4.0 equiv) and [Pd(PPh<sub>3</sub>)<sub>4</sub>] (cat.) in THF/H<sub>2</sub>O (4:1) at ambient temperature<sup>[12]</sup> and 0.01<sub>M</sub> concentration, to afford, much to our surprise, only the monomeric product 29 (72 % yield over the two steps). Increasing the concentration (up to 1.0 m) did not have much effect on the outcome of this reaction, suggesting that the precursor had a well-preorganized disposition towards cyclization once the palladium species was inserted. However, the use of 300 equivalents of TlOEt produced, in addition to 29, the dimeric product 1 in approximately 2% yield (see Scheme 6), after global desilylation. Fluorideinduced global desilylation (TBAF) of 29 (see Scheme 7) gave the all-trans 22-membered ring **m-1** (mono-marinomycin A) and the all-trans 24-membered ring m-2 (iso-mono-marinomycin A), where the lactone had shifted during desilylation, in 85% yield as a separable 1:1 mixture. The monomarinomycins A (m-1 and m-2) were isolated and characterized in pure form by preparative plate chromatography (silica, CH<sub>2</sub>Cl<sub>2</sub>/MeOH 93:7, two elutions) in the dark, followed by HPLC (C18-Dynamax column, 60 Å, 10 mm × 250 mm, 45 % MeCN in H<sub>2</sub>O). iso-Mono-marinomycin A (m-2) yielded crystals (mp: 213°C (dec.), CDCl<sub>3</sub>-D<sub>3</sub>COD) that were suitable for X-ray crystallographic analysis (see ORTEP representation, Figure 1), [13] which confirmed its assigned structure as well as that of its sibling, mono-marinomycin A (m-1) and their precursors. The NOESY, ROESY, and COSY NMR data were also consistent with the assigned structures of m-1 (Table 1) and m-2.

Having realized the propensity of dienyl bromide boronic acid 4 to cyclize before dimerization,

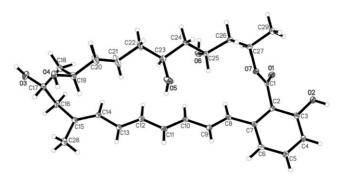


Figure 1. ORTEP representation of iso-mono-marinomycin A (m-2) with thermal ellipsoids shown at the 30% probability level. Hydrogen atoms are shown as white spheres.

we resorted to a stepwise approach to marinomycins A-C. The same key building blocks, 7 (Scheme 5), 26 (Scheme 6), and 28 (Scheme 6) were required, and the revised strategy is continued in Scheme 6. Thus, regioselective hydroboration of hydroxy envne 26 with catecholborane catalyzed by dicyclohexylborane gave the corresponding boronic acid, whose Suzuki coupling (KOH (10 equiv), [Pd(PPh<sub>3</sub>)<sub>4</sub>] (cat.), THF/  $H_2O(4:1)$ ) with dienyl bromide **28** (0.67 equiv) led to hydroxy polyene 30 (63% yield based on 28). Mitsunobu reaction of the latter compound, 30, with carboxylic acid 7 proceeded with inversion of configuration at C25' to afford, after exchange of the Ac group for a TIPS group, enyne ester 31 (78% overall yield for the three steps; Table 1).

Reaction of enyne 31 with catecholborane under the catalytic influence of dicyclohexylborane (THF, 25°C) furnished, after H<sub>2</sub>O quench, the corresponding boronic acid, which was, without isolation, treated with [Pd(PPh<sub>3</sub>)<sub>4</sub>] (stoichiometric) and TIOEt (300 equiv) at ambient temperature to afford the fully protected macrocycle 32. The product was used without purification in the next step, which involved global deprotection of **32** to give marinomycin A (**1**) in 23 % overall yield for the three steps from 31. Synthetic marinomycin A (1) was purified by HPLC (C8-Luna 5μ column,  $100 \text{ Å}, 250 \text{ mm} \times 10 \text{ mm}, 60 \% \text{ MeCN in H}_2\text{O})$  and exhibited identical physical properties ( $R_{\rm f}$ ,  $R_{\rm t}$ , UV spectral,  $\alpha_{\rm D}^{25}$ , mass

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Scheme 6. Preparation of Suzuki coupling precursor 28 and completion of the total synthesis of marinomycins A–C (1–3). Reagents and conditions: a) 5 (1.0 equiv), 6 (1.1 equiv), Ba(OH)<sub>2</sub>·H<sub>2</sub>O (0.75 equiv), THF/H<sub>2</sub>O (20:1), 25 °C, 1 h, 95 %; b) Et<sub>2</sub>BOMe (1.0 м in THF, 1.1 equiv), NaBH<sub>4</sub> (1.1 equiv), THF/MeOH (4:1), -78 °C, 3 h, 89%; c) TBSCI (4.0 equiv), imid. (8.0 equiv), DMF, 25 °C, 8 h, 89%; d) 23 in THF/iPrOH (3:1); then liq. NH<sub>3</sub>; then Ca (30 equiv), -78 °C, 1 h, 75%; e) DMP (1.6 equiv), NaHCO<sub>3</sub> (10 equiv), CH<sub>2</sub>Cl<sub>2</sub>, 25 °C, 30 min, 87%; f) iPr<sub>2</sub>NH (1.8 equiv), nBuLi (2.5 м in hexanes, 1.5 equiv), THF,  $-78 \rightarrow 0$  °C, 30 min, TMSCH<sub>2</sub>N<sub>2</sub> (1.5 equiv), THF, -78 °C, 30 min; then 25, -78 °C, 1 h,  $-78 \rightarrow 25$  °C, 2 h, 85%; g) PPTS (0.1 equiv), EtOH, 25 °C, 3 h, 77%; h) DEAD (6.0 equiv), PPh<sub>3</sub> (6.0 equiv), 7 (6.0 equiv), THF, 25 °C, 1 h, 93 %; i) K<sub>2</sub>CO<sub>3</sub> (0.05 equiv), THF/MeOH (1:1), 25 °C, 15 min; j) TIPSOTf (30 equiv), 2,6-lut. (60 equiv), CH<sub>2</sub>Cl<sub>2</sub>, 25 °C, 18 h, 92% over two steps; k) catecholborane (3.0 equiv), Cy<sub>2</sub>BH (0.1 м in THF, 0.2 equiv), THF, 25 °C, 1 h; then H<sub>2</sub>O (5.0 equiv), 7 (6.0 equiv), THF, 25 °C, 1 h; n) K<sub>2</sub>CO<sub>3</sub> (0.05 equiv), THF/MeOH (1:1), 25 °C, 15 min; o) TIPSOTf (30 equiv), 2,6-lut. (60 equiv), 7 (6.0 equiv), THF, 25 °C, 1 h; n) K<sub>2</sub>CO<sub>3</sub> (0.05 equiv), THF/MeOH (1:1), 25 °C, 15 min; o) TIPSOTf (30 equiv), 2,6-lut. (60 equiv), PPh<sub>3</sub> (6.0 equiv), THF, 25 °C, 1 h; n) K<sub>2</sub>CO<sub>3</sub> (0.05 equiv), THF/MeOH (1:1), 25 °C, 15 min; o) TIPSOTf (30 equiv), 2,6-lut. (60 equiv), CH<sub>2</sub>Cl<sub>2</sub>, 25 °C, 18 h, 78% over three steps; p) catecholborane (3.0 equiv), Cy<sub>2</sub>BH (0.1 м in THF, 0.2 equiv), THF, 25 °C, 1 h; then H<sub>2</sub>O (5.0 equiv); q) TIOEt (300 equiv), [Pd(PPh<sub>3</sub>)<sub>4</sub>] (1.0 equiv), THF/H<sub>2</sub>O (10:1), 25 °C, 4 h; r) TBAF (50 equiv), THF, 25 °C, 18 h, 23% over three steps. Cy=cyclohexyl, DMP=Dess-Martin periodinane, DEAD=diethyl azodicarboxylate, LDA=lithium diisopropylamide, lut.=lutidine, PPTS=pyridinium p-toluenesulfonate, TBAF=tetra-n-butylammonium fluoride, Tf=trifluoromethanesulfonyl

Table 1: Selected physical properties for compounds m-1 and 31.

m-1:  $R_f = 0.54$  (silica gel, CHCl<sub>3</sub>/MeOH/H<sub>2</sub>O 40:9:1);  $[\alpha]_D^{37} = -276.6$  $(c = 0.06, CDCl_3)$ ; IR (film):  $\tilde{v}_{max} = 3352, 2926, 2851, 1709, 1656, 1595,$ 1449, 1376, 1259, 1218, 1118, 1065, 999 cm<sup>-1</sup>; <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>):  $\delta = 7.35$  (t, J = 7.8 Hz, 1 H), 7.13 (d, J = 14.4 Hz, 1 H), 6.92 (d, J = 8.4 Hz, 1 H), 6.77 (d, J = 7.2 Hz, 1 H), 6.47 (dd, J = 14.4, 11.4 Hz, 1 H), 6.34 (t, J = 11.4 Hz, 1 H), 6.16–6.03 (m, 3 H), 5.91 (d, J = 10.8 Hz, 1 H), 5.38-5.20 (m, 3 H); 4.16 (br, 1 H, OH), 4.06 (m, 1 H), 3.95 (m, 1 H), 3.71 (t, J = 9.0 Hz, 1 H), 3.65 (m, 1 H), 3.58 (br, 1 H, OH), 3.29 (br, 1 H, OH), 2.56 (d, J = 10.8 Hz, 1 H), 2.19 (d, J = 13.8 Hz, 1 H), 2.14 (t, J = 10.8 Hz, 1 H, 1.97 - 1.85 (m, 4 H), 1.88 (s, 3 H), 1.79 - 1.75 (m, 2 H),1.70–1.62 (m, 3 H), 1.20 ppm (d, J = 6.0 Hz, 3 H); <sup>13</sup>C NMR (150 MHz,  $CDCl_3$ ):  $\delta = 171.9$ , 162.4, 141.7, 135.8, 135.3, 134.6, 134.5, 133.3, 132.8, 131.9, 131.1, 130.3, 129.7, 128.2, 120.1, 117.0, 73.1, 72.5, 66.4, 65.8, 65.6, 50.4, 48.3, 45.5, 43.8, 42.5, 23.4, 17.8, 14.2 ppm; HRMS (ESI-TOF) calcd for C<sub>29</sub>H<sub>38</sub>O<sub>7</sub>Na [M+Na<sup>+</sup>]: 521.2510; found: 521.2530.

**31**:  $R_f = 0.57$  (silica gel, hexanes/EtOAc, 8:1);  $[\alpha]_D^{37} = -98.8$  (c = 0.09,  $CH_2Cl_2$ ); IR (film):  $\tilde{v}_{max} = 2952$ , 2828, 2856, 1729, 1570, 1465, 1255, 835, 775 cm<sup>-1</sup>; <sup>1</sup>H NMR (600 MHz, C<sub>6</sub>D<sub>6</sub>, mixture of two rotamers around the aryl-carbonyl bond, ca. 1:1 ratio):  $\delta = 7.15$  (J = 8.4 Hz, 1 H), 7.11 (d, J = 8.4 Hz, 1 H), 7.03-6.96 (m, 2 H), 6.94-6.89 (m, 3 H), 6.85 (dd, J = 15.6, 10.8 Hz, 1 H), 6.75 (d, J = 7.8 Hz, 1 H), 6.72 (d, J = 7.8 Hz, 1 H), 6.61 (dd, J = 13.2, 13.2 Hz, 1 H), 6.55 (dd, J = 13.8, 11.4 Hz, 1 H), 6.49– 6.37 (m, 2 H), 6.31 (dd, J = 15.6, 10.8 Hz, 1 H), 6.16 (d, J = 11.4 Hz, 1 H), 6.03 (d, J = 13.8 Hz, 1 H), 5.85 (dt, J = 15.0, 7.2 Hz, 1 H), 5.83 (dt, J = 15.0, 7.2 Hz, 1 H), 5.66 (dd, J = 15.6, 7.2 Hz, 1 H), 5.59 (dd, J = 15.6, 7.2 Hz, 1 H), 5.51–5.45 (m, 2 H), 5.46 (s, 1 H), 4.38 (q, J = 7.2 Hz, 1 H), 4.28 (q, J = 7.2 Hz, 1 H), 4.16–4.05 (m, 6 H), 2.83 (d, J = 2.4 Hz, 1 H), 2.49-2.40 (m, 4 H), 2.33-2.25 (m, 2 H), 2.23-2.10 (m, 6 H), 2.05-1.92 (m, 5 H), 2.04 (s, 3 H), 1.86 (s, 3 H), 1.85-1.79 (m, 2 H), 1.70 (ddd, J = 13.2, 7.2, 4.8 Hz, 1 H, 1.35-1.28 (m, 12 H), 1.19-1.16 (m, 36 H), 1.07(s, 9 H), 1.06 (s, 9 H), 1.06 (s, 27 H), 1.05 (s, 9 H), 1.02 (s, 9 H), 1.01 (s, 9 H), 0.28 (s, 3 H), 0.26 (s, 6 H), 0.23 (s, 3 H), 0.21 (s, 3 H), 0.20 (s, 3 H), 0.19 (s, 3 H), 0.19 (s, 3 H), 0.19 (s, 3 H), 0.16 (s, 6 H), 0.15 (s, 3 H), 0.15 (s, 3 H), 0.14 (s, 3 H), 0.14 (s, 3 H), 0.12 ppm (s, 3 H); <sup>13</sup>C NMR (150 MHz, C<sub>6</sub>D<sub>6</sub>, mixture of rotamers around the aryl-carbonyl bond, ca. 1:1 ratio):  $\delta = 167.4$  (0.5 C), 167.4 (0.5 C) 167.1, 153.9, 153.8 (0.5 C), 153.8 (0.5 C), 150.8, 137.8 (3 C), 137.7, 137.2, 137.1, 136.4, 136.2 (0.5 C), 136.2 (0.5 C), 135.7, 132.4 (0.5 C), 132.3 (0.5 C), 131.5 (0.5 C), 130.9 (0.5 C), 130.9, 130.1, 130.1 (0.5 C), 130.0, 129.9 (0.5 C), 129.1, 129.0, 126.5 (0.5 C), 126.4 (0.5 C), 126.3 (0.5 C), 126.2 (0.5 C), 118.1, 117.9, 117.9, 117.4, 110.6, 110.4 (0.5 C), 110.3 (0.5 C), 107.8, 81.9, 80.7, 72.0, 71.9 (0.5 C), 71.8 (0.5 C), 71.5, 71.4, 69.8 (0.5 C), 69.7 (0.5 C), 68.7, 68.2, 66.5, 66.4 (2 C), 51.7, 48.6, 47.4, 47.2, 46.7, 45.6, 45.5, 42.6, 42.5, 41.0, 30.2, 26.3 (9 C), 26.2 (9 C), 26.2 (3 C), 26.2 (3 C), 24.5, 24.4, 20.3, 18.3 (8C), 17.9 (12C), 12.7 (6C), -3.4, -3.4, -3.8 (2C), -3.9 (3C), -3.9, -4.0, -4.1 (4 C), -4.2, -4.3, -4.4 ppm; HRMS (ESI-TOF) calcd for  $C_{124}H_{226}BrO_{14}Si_{10}$  [M-H<sup>-</sup>]: 2298.3853; found: 2298.3852.

spectrometric, and <sup>1</sup>H and <sup>13</sup>C NMR spectral data) to those recorded for the naturally occurring substance. [1,14] When allowed to isomerize in ambient light, marinomycin A (1) formed mixtures with marinomycins B (2) and C (3) as previously reported ( $1/2/3 \approx 16:2:9$  after 30 min; ca. 1:1:1 after 2 h by HPLC).[1,15]

The success of the Suzuki reaction in these syntheses underscores its usefulness in the construction of complex molecules. Besides rendering the naturally occurring marinomycins A-C (1-3) readily available, the described synthetic technology also provides access to their monomeric derivatives, mono-marinomycin A (m-1) and its isomer iso-mono-

Scheme 7. Formation of mono-marinomycins m-1 and m-2. Reagents and conditions: a) catecholborane (3.0 equiv), Cy<sub>2</sub>BH (0.1 M in THF, 0.2 equiv), THF, 25 °C, 1 h; then H<sub>2</sub>O (5.0 equiv); b) TIOEt (4.0 equiv), [Pd(PPh<sub>3</sub>)<sub>4</sub>] (0.1 equiv), THF/H<sub>2</sub>O (4:1), 25 °C, 30 min, 72 % over two steps; c) TBAF (30 equiv), THF, 18 h, 85 % yield.

marinomycin A (m-2), and opens the way to the construction of other members of the class of natural or designed origins.

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- [15] Exact ratios of marinomycins A–C (1–3) varied from one experiment to another.